

Appendix 1: Implementation in the R software

The likelihood for an ordinary spatial capture-recapture model in the absence of class structure is specified by the R function `intlik2` from Royle et al. (2014):

```
intlik2 <- function(parm,y=y,X=traplocs,delta=.3,ssbuffer=2){

  Xl <- min(X[,1]) - ssbuffer
  Xu <- max(X[,1]) + ssbuffer
  Yu <- max(X[,2]) + ssbuffer
  Yl <- min(X[,2]) - ssbuffer

  xg <- seq(Xl+delta/2,Xu-delta/2,delta)
  yg <- seq(Yl+delta/2,Yu-delta/2,delta)
  npix.x <- length(xg)
  npix.y <- plength(yg)

  G <- cbind(rep(xg,npix.y),sort(rep(yg,npix.x)))
  nG <- nrow(G)
  D <- e2dist(X,G)
  area<- (delta*delta)*nrow(G)

  # extract the parameters from the input vector
  alpha0 <- parm[1]
  alpha1 <- exp(parm[2])
  n0 <- exp(parm[3]) # note parm[3] lives on the real line
  probcap <- plogis(alpha0)*exp(-alpha1*D*D)
  Pm <- matrix(NA,nrow=nrow(probcap),ncol=ncol(probcap))
  ymat <- rbind(y,rep(0,ncol(y))) ## tack on all-zero history
  lik.marg <- rep(NA,nrow(ymat))
  for(i in 1:nrow(ymat)){
    Pm[1:length(Pm)] <- dbinom(rep(ymat[i,],nG), K,
                              probcap[1:length(Pm)], log=TRUE))
    lik.cond <- exp(colSums(Pm))
    lik.marg[i] <- sum( lik.cond*(1/nG) )
  }
  nv <- c(rep(1,length(lik.marg)-1),n0)
  ## part1 here is the combinatorial term,
  ## log(factorial(N)) = lgamma(N+1)
  part1 <- lgamma(nrow(y)+n0+1) - lgamma(n0+1)
  part2 <- sum(nv*log(lik.marg))
  return( -1*(part1+ part2) )
}
```

This function uses an encounter probability model that is proportional to the kernel of a Gaussian probability distribution function (“half-normal” in distance sampling),

$$p(\mathbf{x}, \mathbf{s}) = \text{logit}^{-1}(\alpha_0) \exp(-\alpha_1 \|\mathbf{x} - \mathbf{s}\|^2)$$

where $\text{logit}^{-1}(\alpha_0)$ is the baseline encounter probability in trap \mathbf{x} , $\|\mathbf{x} - \mathbf{s}\|$ is the distance between trap \mathbf{x} and activity center \mathbf{s} , and $\alpha_1 = 1/(2\sigma^2)$. In the optimization of the likelihood we estimate the parameters α_0 and $\log(\alpha_1)$ for numerical stability.

The function takes an argument, the unknown parameters of the model, and additional arguments including the encounter history matrix \mathbf{y} , the trap locations `traplocs`, the spacing of the integration grid (argument `delta`) and the state-space buffer. The first few lines of the function compute a grid of points used to evaluate the integrand, and the remaining half of the function computes the marginal likelihood contributions for each individual and the “all zero” encounter history. R code for simulating data and optimizing the likelihood using this function is in the `scrbook` R package from Royle et al. (2014) which can be viewed by looking at the help file `?intlrik2`.

In the following R script, we provide some functions for simulating data for a 2-class situation (e.g., sex structured models) and a modification of the `intlrik2` function to fit models with or without sex. The function `simSCR0sex` simulates sex-structured data along

with a given fraction of missing data (argument `na.frac`). The simulation function also simulates data with a behavioral response. The more general likelihood function is provided in the R package `scrbook` (Royle et al. 2014) available from the authors.

```
### This function simulates data with a sex-specific
### intercept and some missing sex values (na.frac), also
### allows for a behavioural response (beta).

e2dist<- function (x, y){
  i <- sort(rep(1:nrow(y), nrow(x)))
  dvec <- sqrt((x[, 1] - y[i, 1])^2 + (x[, 2] - y[i, 2])^2)
  matrix(dvec, nrow = nrow(x), ncol = nrow(y), byrow = F)
}

simSCR0sex <- function (N = 100, K = 20, alpha0 = c(-2.5,
  -1.8), sigma = c(0.5,0.75), beta=0, discard0 = TRUE,
  array3d = FALSE, psi.sex=.35,theta = .9){
  ### beta = behavioral response
  ### theta = PER SAMPLE probability of obtaining sex info
  traplocs <- cbind(sort(rep(1:7, 7)), rep(1:7, 7))
  Dmat <- e2dist(traplocs, traplocs)
  ntraps <- nrow(traplocs)
  plot(traplocs)
  buffer <- 2
  Xl <- min(traplocs[, 1] - buffer)
  Xu <- max(traplocs[, 1] + buffer)
  Yl <- min(traplocs[, 2] - buffer)
  Yu <- max(traplocs[, 2] + buffer)
  sx <- runif(N, Xl, Xu)
  sy <- runif(N, Yl, Yu)
  sex<- rbinom(N,1,psi.sex)
  if(length(alpha0) != 2){
    alpha0<- c(alpha0,alpha0)
  }
  S <- cbind(sx, sy)
  D <- e2dist(S, traplocs)
  if(length(sigma) != 2){
    sigma<- c(sigma, sigma)
  }

  alpha1 <- 1/(2 * sigma * sigma)
  probcap1 <- plogis(alpha0[1])*exp(-alpha1[1] * D * D)
  probcap2 <- plogis(alpha0[2])*exp(-alpha1[2] * D * D)
```

```

Y <- matrix(NA, nrow = N, ncol = ntraps)
for(i in 1:nrow(Y)){
  if(sex[i]==0)
    Y[i, ] <- rbinom(ntraps, K, probcap1[i, ])
  if(sex[i]==1)
    Y[i, ] <- rbinom(ntraps, K, probcap2[i, ])
}
if(discard0) {
  ncaps <- apply(Y, 1, sum)
  Y <- Y[ncaps > 0, ]
}
dimnames(Y)<- list(1:nrow(Y), paste("trap", 1:ncol(Y), sep=""))
if(array3d){
  Y <- array(NA, dim = c(N, ntraps, K))
  for (i in 1:nrow(Y)) {
    for (j in 1:ntraps) {
      prevcap<- 0
      Y[i,j,1]<- rbinom(1,1,plogis(alpha0[sex[i]+1] )*exp(-
alpha1[sex[i]+1]*D[i,j]*D[i,j]) )
      for(k in 2:K){
        prevcap<- sum(Y[i,j,1:(k-1)])>0
        Y[i,j,k]<- rbinom(1,1,plogis(alpha0[sex[i]+1] + beta*prevcap)*exp(-
alpha1[sex[i]+1]*D[i,j]*D[i,j]) )
      }
    }
  }
  if (discard0) {
    Y2d <- apply(Y, c(1, 2), sum)
    ncaps <- apply(Y2d, 1, sum)
    Y <- Y[ncaps > 0, , ]
  }
}
sex.obs<-sex[ncaps>0]

p.obtain.sex<- 1-(1-theta)^ncaps[ncaps>0]
obtain.sex<- rbinom(length(sex.obs),1,p.obtain.sex)
sex.obs[!obtain.sex]<- NA
list(Y = Y, traplocs = traplocs, xlim = c(Xl, Xu), ylim = c(Yl,
Yu), N = N, alpha0 = alpha0, alpha1 = alpha1, sigma = sigma,
K = K,sex=sex.obs)
}

```

```

### Here is a likelihood function patterned off of intlik2
### from the "scrbook" package. Takes argument "sex" (observed data)
### parm should be 5 in length:
### alpha0[1] alpha0[2], log(alpha1), log(n0), logit(psi.sex)

```

```

intlik2sex<-
function (parm, y = y, delta = 0.3, K, X = traplocs, sex, ssbuffer =
2,sex.int=TRUE,sex.sig=TRUE)
{
  sex<-sex+1
  Xl <- min(X[, 1]) - ssbuffer
  Xu <- max(X[, 1]) + ssbuffer
  Yu <- max(X[, 2]) + ssbuffer
  Yl <- min(X[, 2]) - ssbuffer
  xg <- seq(Xl + delta/2, Xu - delta/2, delta)
  yg <- seq(Yl + delta/2, Yu - delta/2, delta)
  npix.x <- length(xg)
  npix.y <- length(yg)

  G <- cbind(rep(xg, npix.y), sort(rep(yg, npix.x)))
  nG <- nrow(G)
  D <- e2dist(X, G)

  area<- (delta*delta)*nrow(G)

  if(!sex.sig & sex.int){
    alpha0<- parm[1:2]
    alpha1 <- c(exp(parm[3]),exp(parm[3]))
    n0 <- exp(parm[4])
    psi.sex<-plogis(parm[5])
  }

  if(sex.sig & sex.int){
    alpha0<- parm[1:2]
    alpha1 <- exp(parm[3:4])
    n0 <- exp(parm[5])
    psi.sex<-plogis(parm[6])
  }
  if(sex.sig & !sex.int){
    alpha0<- c(parm[1],parm[1])
    alpha1<- exp(parm[2:3])
    n0 <- exp(parm[4])
    psi.sex<-plogis(parm[5])
  }
  if(!sex.sig & !sex.int){
    alpha0<- c(parm[1],parm[1])
    alpha1<- c(exp(parm[2]),exp(parm[2]))
    n0 <- exp(parm[3])
    psi.sex<-plogis(parm[4])
  }

  probcap1 <- plogis(alpha0[1]) * exp(-alpha1[1] * D * D)
  probcap2 <- plogis(alpha0[2]) * exp(-alpha1[2] * D * D)

```

```

Pm1<-Pm2 <- matrix(NA, nrow = nrow(probcap1), ncol = ncol(probcap1))
ymat <- y
ymat <- rbind(y, rep(0, ncol(y)))
lik.marg1<-lik.marg2 <- rep(NA, nrow(ymat))
lik.marg<- lik.marg1
for (i in 1:nrow(ymat)) {

    Pm1[1:length(Pm1)] <- (dbinom(rep(ymat[i, ], nG), rep(K,nG),
probcap1[1:length(Pm1)],
    log = TRUE))
    lik.cond1 <- exp(colSums(Pm1))
    lik.marg1[i] <- sum(lik.cond1 * (1/nG))

    Pm2[1:length(Pm2)] <- (dbinom(rep(ymat[i, ], nG), rep(K,nG),
probcap2[1:length(Pm2)],
    log = TRUE))
    lik.cond2 <- exp(colSums(Pm2))
    lik.marg2[i] <- sum(lik.cond2 * (1/nG))

    if (sex[i]==1 & !is.na(sex[i]))
    lik.marg[i]<- lik.marg1[i]*(1-psi.sex)

    if (sex[i]==2 & !is.na(sex[i]))
    lik.marg[i]<- lik.marg2[i]*psi.sex

    if (is.na(sex[i]))
    lik.marg[i]<- lik.marg1[i]*(1-psi.sex) + lik.marg2[i]*psi.sex

}

nv <- c(rep(1, length(lik.marg) - 1), n0)
part1 <- lgamma(nrow(y) + n0 + 1) - lgamma(n0 + 1)
part2 <- sum(nv * log(lik.marg))
-1 * (part1 + part2)
}

```

The following set of R commands simulate a data set and fit the sex-structured model. The nature of the sex-specificity of model parameters is determined by the logical flags `sex.int` and `sex.sig`. If one or the other are TRUE then either or both the intercept or σ , respectively, are estimated as sex-specific parameters.

```

## Simulate a data set with no missing sex values and
## sex-specific intercepts but NOT sex-specific sigma
## Note "rnd" is the random number seed.

```

```

data<-simSCR0sex(N=200,discard0=TRUE,alpha0=c(-2.5,-1.8),
sigma=c(.75,.75),beta=0,theta=1,psi.sex=0.50)

## Harvest the elements of the data set
y<-data$Y
traplocs<-data$traplocs
nind<-nrow(y)
J<-nrow(traplocs)
K<-data$K
sex<-data$sex

##
## First we obtain the MLEs for a model with sex-specific intercept
## but not sex-specific sigma.

frog1<-nlm(intlik2sex,c(-2.5,-2,1.0,log(4),0),
hessian=TRUE,y=y,X=traplocs,K=rep(K,J),
sex=sex,delta=.3,ssbuffer=2,sex.int=TRUE,sex.sig=FALSE)

##
## Next we obtain MLEs for a model with NO sex-specificity but we
## still estimate the probability of sex = 1

frog2<-nlm(intlik2sex,c(-2.5,1.5,log(4),0),hessian=TRUE,y=y,
X=traplocs,K=rep(K,J),
sex=sex,delta=.3,ssbuffer=2,sex.int=FALSE,sex.sig=FALSE)

```

